

# 成纤维细胞生长因子19/15的表达调节与生物活性

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## 摘要

FGF19(啮齿动物中FGF15)是内分泌FGF家族的成员之一,属于肠道激素,参与调节胆汁酸、葡萄糖和脂质代谢等,维持全身稳态。FGF19/15对多种慢性疾病具有治疗潜力,包括肥胖症、糖尿病和恶性肿瘤等。本文总结FGF19/15的表达调节及药理活性的研究进展,初步评估其临床应用潜力。

## 关键词

FGF19/15, 胆汁酸稳态, 糖脂代谢

# Expression Regulation and Bioactivity of Fibroblast Growth Factor 19/15

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## Abstract

FGF19 (FGF15 in rodents), a member of the endocrine FGF family, is an intestinal hormone involved in the regulation of bile acid, glucose and lipid metabolism to maintain systemic homeostasis. FGF19/15 has the potential to treat a variety of chronic diseases, including obesity, diabetes and malignant cancers. This paper reviews the biological activities of FGF19/15, and preliminarily evaluates the clinical application.

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## Keywords

FGF19/15, Bile Acid Homeostasis, Metabolism of Glucose and Lipid

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## 1. 引言

1999年,成纤维细胞生长因子19(Fibroblast growth factor, FGF19)在人脑中首次被发现,其编码基因位于染色体11q13,由216个氨基酸组成,在回肠和胆囊上皮细胞中高表达[1],但在正常肝脏中检测不到[2]。成年小鼠的回肠、空肠和十二指肠中高度表达FGF15,是FGF19的小鼠同源物,由218个氨基酸组成,与人FGF19具有51%的相似性。与其他具有旁分泌和自分泌功能的FGF不同,FGF19/15对硫酸乙酰肝素蛋白聚糖(HSPG)的亲合力低,这使得它们在细胞中合成后分泌进入细胞外基质,并且作为内分泌激素参与循环[3]。除了在调节胆汁酸稳态中的核心作用外,FGF19对代谢性疾病(如肥胖和糖尿病)亦具有治疗潜力[4]。

本文总结FGF19/15表达的调控信号及生物活性,对其潜在的临床应用前景进行分析。

## 2. FGF19/15表达的调控因素及信号传导

FGF19基因的启动子区含有功能性法尼醇X受体反应元件(FXRE),当胆汁酸作用于法尼醇X受体(FXR)后,FXR与FXRE结合,诱导FGF19/15在回肠上皮细胞中表达。最近的研究表明,FGF19/15表达还受到其他成分错综复杂的调节,如:1)甾醇调节元件结合蛋白2(SREBP2)通过抑制FXR与FGF19启动子中的FXRE结合,负性调控肠癌细胞中FGF19的转录[5];2)核受体维生素D受体(VDR)[6]、类视黄醇X受体(RXR)和孕烷X受体(PXR)[7]等均可诱导小鼠肠道中FGF15表达;3)新发现的肠道蛋白包含的甲基多巴-A5-酪氨酸磷酸酶(MAM)和低密度脂蛋白(LDL)受体A类结构域1(MALRD1)能促进FGF19/15转录[8];4)Wnt调节转录因子TCF7L2基因的转录也能上调小鼠肠道中FGF15表达水平[9]。

分泌的FGF19/15与首选受体FGFR4和辅助受体 $\beta$ -klotho结合[10],募集细胞溶质接头蛋白(如支架蛋白—成纤维细胞生长因子受体底物2 $\alpha$ (FRS2 $\alpha$ )),从而触发信号级联反应,呈现抑制肝胆汁酸合成等作用。虽然肝脏是FGF19激活FGFR4- $\beta$ -klotho复合物信号传导的主要部位,但FGF19在其他组织中也有多重生物学功能,在白色脂肪组织(WAT),FGF19结合并激活其中的FGFR1c- $\beta$ -klotho[11]。细胞外信号调节激酶1(ERK1)、ERK2和原癌基因Ras-ERK-p90等下游信号可介导FGF19-FGFR4- $\beta$ -klotho复合物的作用[12][13]。

## 3. FGF19/15的生物活性

### 3.1. 调控胆汁酸稳态

胆汁酸是肝脏中胆固醇分解代谢产生的强清洁剂,具有强乳化作用和毒性,其合成须受到严格调控。餐后胆囊收缩,将胆汁酸释放到肠道中,协助脂质和脂溶性维生素溶解和吸收[14][15]。FXR刺激FGF19/15在小肠中表达,后者通过靶向FGFR4和孤核受体小异源二聚体伴侣(SHP)抑制肝脏中的胆固醇7 $\alpha$ -羟化酶(CYP7A1)活性和表达,该酶负责催化经典胆汁酸合成途径中的第一步和限速步骤。

在化学诱导的胆汁淤积性肝损伤模型小鼠回肠中 FGF15 表达降低[16]。缺乏 FGF15 的小鼠肝脏中 CYP7A1 mRNA 和蛋白水平增加, CYP7A1 酶活性和粪便胆汁酸排泄相应增加。Fgf15<sup>-/-</sup>、Fgfr4<sup>-/-</sup>和 $\beta$ -klotho<sup>-/-</sup>敲除动物的胆汁酸代谢失调, 而在 Fgfr4<sup>-/-</sup>和 $\beta$ -klotho<sup>-/-</sup>敲除动物中使用人源 FGF19 未能抑制 CYP7A1 表达[11]。而给予 FGF19 能够降低人原代肝细胞中 CYP7A1 的 mRNA 表达水平, 给予胆汁酸或 FXR 激动剂诱导小鼠回肠上皮细胞表达 FGF15, 能够抑制肝脏中 CYP7A1 表达, 阻止胆汁酸的合成[17]。

NGM282 是 FGF19 合成衍生物, 其对肝脏疾病尤其是胆汁淤积性肝病具有改善作用[18]。患有胆汁淤积性肝病的患者使用 NGM282 后, 降低了具有高细胞毒性和去污活性的甘氨酸结合型疏水性胆汁酸表达水平, 这为 NGM282 治疗胃肠道和肝脏疾病提供了可能[19]。此外, FGF19 类似物的辅助治疗也能够改善对熊去氧胆酸(UDCA)治疗无反应的原发性胆汁型肝硬化患者的疾病症状[4]。

## 3.2. 调节糖脂代谢

### 3.2.1. 对糖代谢的影响

除了调节胆汁酸稳态外, FGF19/15 参与维持糖代谢[20], 代谢综合征或二型糖尿病患者的循环 FGF19 水平低于健康对照组[21]。Fgf15 基因敲除小鼠不能正常维持血液中的葡萄糖浓度, 并表现出肝糖原水平降低和葡萄糖耐受不良的现象, 而这些可通过外源性给予 FGF19 的方式得到改善[12], 而给予 Fgfr4 敲除小鼠 FGF15 并不能起到类似作用。

腺相关病毒(AAV)过表达 FGF15 可增加饮食诱导的肥胖(DIO)小鼠的能量消耗, 限制 db/db 小鼠糖尿病的发生[22]。在胰岛素抵抗的小鼠模型中, 脑室内注射 FGF19 可改善血糖状态并增强外周胰岛素信号传导, 改善糖尿病大鼠的葡萄糖代谢[23]。静脉注射 FGF19 可降低 DIO 和 ob/ob 小鼠的体重, 提高葡萄糖利用率[24]。机制上 FGF19/15 通过抑制环磷酸腺苷(cAMP)反应元件结合蛋白-过氧化物酶体增殖物激活受体- $\gamma$  共激活剂-1 $\alpha$  (CREB-PGC-1 $\alpha$ )信号传导, 抑制葡萄糖的肝脏代谢[25]。

### 3.2.2. 对脂质代谢的影响

FGF19/15 在调节脂质代谢中同样起着重要作用。肥胖患者血液中的 FGF19 浓度低于健康受试者[4][26], 非酒精性脂肪肝病(NASH)患者血清的 FGF19 浓度降低[27]。

Fgf15 敲除降低 HFD 诱导的小鼠血清甘油三酯增加以及脂质代谢酶基因的表达, 表明 FGF15 改善了小鼠脂质代谢[28]。人源 FGF19 可降低小鼠体重, 防止小鼠脂肪沉积, 降低丙氨酸氨基转移酶(ALT)活性和肝脏脂质(如甘油三酯和游离脂肪酸)水平, 增加肝脏脂肪酸氧化[29]。长效的 FGF19/载脂蛋白嵌合分子治疗可减少小鼠体内肝脏脂质积累[30], 证明 FGF19 及其类似物可改善代谢性疾病模型非酒精性脂肪性肝炎(NASH)的病理学特征, 呈现抗脂质蓄积、抗炎和抗纤维化的作用[31][32]。

目前, NGM282 的 I 期安全性和耐受性研究以及 NASH 患者中为期 12 周的 II 期安全性、耐受性和有效性研究已经完成[33]。II 期临床的研究结果与临床前动物研究的结果大体一致, NGM282 能够降低体重和身体质量指数(BMI)。肝活检的组织学评估发现, 84%的患者疾病活动性评分(NAS 评分)有所改善, 42%的患者的纤维化分期有所改善。II 期研究结果与临床前动物研究的主要区别在于给药后血脂水平差异[34]。24 周后, 接受 NGM282 治疗的 NASH 患者肝脏脂肪含量显著降低, 并伴随纤维化程度的改善[35]。

机制研究发现, FGF19 显著降低与脂肪生成相关基因的转录水平, 包括乙酰辅酶 A 羧化酶(ACC)、血小板糖蛋白 4 (CD36)、转录因子甾醇调节元件结合蛋白-1c (SREBP-1c)、硬脂酰辅酶 A1 (SCD1)和 CYP7A1 [29]。另外, 在 FGFR1c/KLB 敲除小鼠中, FGF19 缺乏代谢活性, 表明 FGF19 可依赖 FGFR1c/KLB 表达降低肝脂肪变性[36], 而肝脏 FGFR4 对于 FGF19 改善高脂肪饮食喂养小鼠的脂质代谢可能并不是必需的[37]。

棕榈酸是一种在动物及植物内普遍存在的脂肪酸, 过量的棕榈酸会抑制葡萄糖摄取, 增加胰岛素抵抗, 上调线粒体  $\beta$ -氧化, 导致线粒体过载, 增加活性氧(ROS)产生。给予 FGF19 可抑制小鼠成肌细胞的

棕榈酸酯  $\beta$ -氧化, 减轻线粒体过载并防止线粒体介导的细胞凋亡[38]。FGF19 促进线粒体中 PGC1 $\alpha$ 、线粒体转录因子 A (TFAM)、血红素加氧酶 1 (HO-1)等表达, 改善棕榈酸诱导的线粒体功能障碍[39]。

### 3.3. 促进有丝分裂作用

FGF15 在小鼠具有促进有丝分裂活性, 能够诱导肝细胞增殖, 在促进肝再生方面至关重要, Fgf15 缺乏的小鼠经历肝切除术之后表现出肝再生受损、明显的肝损伤和更高的死亡率[40], 这种活性导致 FGF19/15 可能与肝细胞性肝癌(HCC)相关。FGF15 的异位表达促进肝细胞增殖、发育异常和肿瘤形成, 这种致癌作用与 FGFR4 密切相关, FGFR4 缺失或使用 FGFR4 中和抗体能够减少小鼠异位 FGF19 表达后的肿瘤负荷[41]。靶向 FGF19-FGFR4 通路可有效抑制临床前动物模型中 HCC 的发展。FGF19 也可能是肺鳞状细胞癌(LSQ)的有效标志物和潜在驱动因素[42], 非小细胞肺癌患者的血清 FGF19 水平明显高于对照组。在 LSQ 的小鼠体内模型中, FGF19 的过表达促进了肿瘤细胞的增殖和迁移[43]。

鉴于 FGF19 的致癌潜能, 基于结构-功能原理对其进行改构十分必要。已有报道通过诱变 FGF19 的五个 N 端和肝素结合区, 分离 FGF19 的有丝分裂活性和代谢活性, 合成了一系列 FGF19 变体, 它们保留了有益的代谢调节作用, 减少了致癌性[44]。

### 3.4. 其他

给予重组 FGF19 蛋白可以升高小鼠骨骼肌质量和强度, 增加人肌肉细胞在从成肌细胞分化为肌管过程中的肌管面积[45]。FGF19 通过上调肌肉细胞生长的关键调节剂核糖体蛋白 S6 激酶(S6K1)的磷酸化水平增强肌肉纤维, 改善小鼠肌肉萎缩的症状[46]。

冠状动脉疾病(CAD)是一种缺血性心脏病, 包括稳定型心绞痛、不稳定型心绞痛、心肌梗死和心源性猝死。血清 FGF19/15 水平与 CAD 的存在和严重程度呈负相关。Fgf15 基因敲除小鼠存在心脏缺陷、主动脉和肺动脉干排列不良等症状, 这种情况与心室流出道早期发育的形态缺陷相关, 表明 FGF19/15 是心室流出道发育所必需的[47]。

## 4. 结语

自发现以来, 几项突破性研究使人们对 FGF19/15 的功能和疾病治疗潜力有了更深入的了解。但基于 FGF19 的疗法面临的主要问题是: 1) 存在潜在的脱靶效应; 2) 天然 FGF19 蛋白分子量较小(约 25 kDa), 通过肾脏后被迅速过滤清除, 导致其半衰期短(约 30 min) [20], 药代动力学性质较差; 3) FGF19/15 具有的致癌潜能[48]。

针对 FGF19/15 成药受阻的原因, 需对其进行有目的性的改构, 同时考虑 FGF19 的药效机制与抑制 FGF19-FGFR4 通路, 分离其调节代谢活性区域与致癌活性区域, 对后者进行结构改变, 避免潜在的不利影响。NGM282 是 FGF19 的非致癌性变体, 尽管其长期治疗的有效性和安全性仍需要进一步验证, 目前认为它是最有希望的基于 FGF19 的内分泌干预手段。

总之, FGF19/15 是调节代谢和器官间信号传导的重要激素, 可作为疾病的危险因素和生物标志物。目前, 靶向 FGF19 疗法(使用 FGF19 类似物或使用 FXR 激动剂)已取得较好的成果, 而针对肠道疾病相关的 FGF19 缺乏症的治疗途径有待开拓。

## 参考文献

- [1] Phan, P., Saikia, B.B., Sonnaila, S., *et al.* (2021) The Saga of Endocrine FGFs. *Cells*, **10**, Article No. 2418. <https://doi.org/10.3390/cells10092418>
- [2] Naugler, E., Tarlow, D., Fedorov, M., *et al.* (2015) Fibroblast Growth Factor Signaling Controls Liver Size in Mice

- with Humanized Livers. *Gastroenterology*, **149**, 728-740. <https://doi.org/10.1053/j.gastro.2015.05.043>
- [3] Itoh, N., Ohta, H. and Konishi, M. (2015) Endocrine FGFs: Evolution, Physiology, Pathophysiology, and Pharmacotherapy. *Frontiers in Endocrinology*, **6**, Article No. 154. <https://doi.org/10.3389/fendo.2015.00154>
- [4] Zhang, F., Yu, L., Lin, X., *et al.* (2015) Minireview: Roles of Fibroblast Growth Factors 19 and 21 in Metabolic Regulation and Chronic Diseases. *Molecular Endocrinology*, **29**, 1400-1413. <https://doi.org/10.1210/me.2015-1155>
- [5] Miyata, M., Hata, T., Yamazoe, Y., *et al.* (2014) SREBP-2 Negatively Regulates FXR-Dependent Transcription of FGF19 in Human Intestinal Cells. *Biochemical and Biophysical Research Communications*, **443**, 477-482. <https://doi.org/10.1016/j.bbrc.2013.11.126>
- [6] Li, J., Witonsky, D., Sprague, E., *et al.* (2021) Genomic and Epigenomic Active Vitamin D Responses in Human Colonic Organoids. *Physiological Genomics*, **53**, 235-248. <https://doi.org/10.1152/physiolgenomics.00150.2020>
- [7] Guthrie, G., Stoll, B., Chacko, S., *et al.* (2020) Rifampicin, Not Vitamin E, Suppresses Parenteral Nutrition-Associated Liver Disease Development through the Pregnane X Receptor Pathway in Piglets. *American Journal of Physiology—Gastrointestinal and Liver Physiology*, **318**, G41-G52. <https://doi.org/10.1152/ajpgi.00193.2019>
- [8] Wang, L., Frey, M. and Kohli, R. (2021) The Role of FGF19 and MALRD1 in Enterohepatic Bile Acid Signaling. *Frontiers in Endocrinology*, **12**, Article ID: 799648. <https://doi.org/10.3389/fendo.2021.799648>
- [9] Bhat, N., Esteghamat, F., Chaube, K., *et al.* (2022) TCF7L2 Transcriptionally Regulates Fgf15 to Maintain Bile Acid and Lipid Homeostasis through Gut-Liver Crosstalk. *The FASEB Journal*, **36**, e22185. <https://doi.org/10.1096/fj.202101607R>
- [10] Somm, E. and Jornayvaz, R. (2018) Fibroblast Growth Factor 15/19: From Basic Functions to Therapeutic Perspectives. *Endocrine Reviews*, **39**, 960-989. <https://doi.org/10.1210/er.2018-00134>
- [11] Tomiyama, K., Maeda, R., Urakawa, I., *et al.* (2010) Relevant Use of Klotho in FGF19 Subfamily Signaling System *in Vivo*. *Proceedings of the National Academy of Sciences of the United States of America*, **107**, 1666-1671. <https://doi.org/10.1073/pnas.0913986107>
- [12] Kir, S., Beddow, S.A., Samuel, V.T., *et al.* (2011) FGF19 as a Postprandial, Insulin-Independent Activator of Hepatic Protein and Glycogen Synthesis. *Science*, **331**, 1621-1624. <https://doi.org/10.1126/science.1198363>
- [13] Kong, B., Wang, L., Chiang, J.Y.L., *et al.* (2012) Mechanism of Tissue-Specific Farnesoid X Receptor in Suppressing the Expression of Genes in Bile-Acid Synthesis in Mice. *Hepatology*, **56**, 1034-1043. <https://doi.org/10.1002/hep.25740>
- [14] Johansson, H., Mörk, L.M., Li, M., *et al.* (2018) Circulating Fibroblast Growth Factor 19 in Portal and Systemic Blood. *Journal of Clinical and Experimental Hepatology*, **8**, 162-168. <https://doi.org/10.1016/j.jceh.2017.07.001>
- [15] Xu, A.W. (2018) Hypothalamic Sensing of Bile Acids, a Gut Feeling. *Trends in Endocrinology & Metabolism*, **29**, 363-366. <https://doi.org/10.1016/j.tem.2018.02.001>
- [16] Tang, X., Yang, Q., Yang, F., *et al.* (2016) Target Profiling Analyses of Bile Acids in the Evaluation of Hepatoprotective Effect of Gentiopicroside on ANIT-Induced Cholestatic Liver Injury in Mice. *Journal of Ethnopharmacology*, **194**, 63-71. <https://doi.org/10.1016/j.jep.2016.08.049>
- [17] Al-Khaifi, A., Rudling, M. and Angelin, B. (2018) An FXR Agonist Reduces Bile acid Synthesis Independently of Increases in FGF19 in Healthy Volunteers. *Gastroenterology*, **155**, 1012-1016. <https://doi.org/10.1053/j.gastro.2018.06.038>
- [18] Luo, J., Ko, B., Elliott, M., *et al.* (2014) A Nontumorigenic Variant of FGF19 Treats Cholestatic Liver Diseases. *Science Translational Medicine*, **6**, ra100. <https://doi.org/10.1126/scitranslmed.3009098>
- [19] Sanyal, J., Ling, L., Beuers, U., *et al.* (2021) Potent Suppression of Hydrophobic Bile Acids by Aldafermin, an FGF19 Analogue, across Metabolic and Cholestatic Liver Diseases. *JHEP Reports*, **3**, Article No. 100255. <https://doi.org/10.1016/j.jhepr.2021.100255>
- [20] Potthoff, J., Kliewer, A. and Mangelsdorf, D.J. (2012) Endocrine Fibroblast Growth Factors 15/19 and 21: From Feast to Famine. *Genes & Development*, **26**, 312-324. <https://doi.org/10.1101/gad.184788.111>
- [21] Sonne, D.P., Van Nierop, F.S., Kulik, W., *et al.* (2016) Postprandial Plasma Concentrations of Individual Bile Acids and FGF-19 in Patients with Type 2 Diabetes. *The Journal of Clinical Endocrinology & Metabolism*, **101**, 3002-3009. <https://doi.org/10.1210/jc.2016-1607>
- [22] Zhou, M., Luo, J., Chen, M., *et al.* (2017) Mouse Species-Specific Control of Hepatocarcinogenesis and Metabolism by FGF19/FGF15. *Journal of Hepatology*, **66**, 1182-1192. <https://doi.org/10.1016/j.jhep.2017.01.027>
- [23] Morton, J., Matsen, E., Bracy, P., *et al.* (2013) FGF19 Action in the Brain Induces Insulin-Independent Glucose Lowering. *Journal of Clinical Investigation*, **123**, 4799-808. <https://doi.org/10.1172/JCI70710>
- [24] Fu, L., John, L.M., Adams, S.H., *et al.* (2004) Fibroblast Growth Factor 19 Increases Metabolic Rate and Reverses Dietary and Leptin-Deficient Diabetes. *Endocrinology*, **145**, 2594-603. <https://doi.org/10.1210/en.2003-1671>

- [25] Potthoff, M.J., Boney-Montoya, J., Choi, M., *et al.* (2011) FGF15/19 Regulates Hepatic Glucose Metabolism by Inhibiting the CREB-PGC-1 $\alpha$  Pathway. *Cell Metabolism*, **13**, 729-738. <https://doi.org/10.1016/j.cmet.2011.03.019>
- [26] Hu, X., Xiong, Q., Xu, Y., *et al.* (2018) Association of Serum Fibroblast Growth Factor 19 Levels with Visceral Fat Accumulation Is Independent of Glucose Tolerance Status. *Nutrition, Metabolism & Cardiovascular Diseases*, **28**, 119-125. <https://doi.org/10.1016/j.numecd.2017.10.009>
- [27] Schreuder, C.M.A., Marsman, H.A., Lenicek, M., *et al.* (2010) The Hepatic Response to FGF19 Is Impaired in Patients with Nonalcoholic Fatty Liver Disease and Insulin Resistance. *American Journal of Physiology—Gastrointestinal and Liver Physiology*, **298**, G440-G445. <https://doi.org/10.1152/ajpgi.00322.2009>
- [28] Schumacher, J.D., Kong, B., Pan, Y., *et al.* (2017) The Effect of Fibroblast Growth Factor 15 Deficiency on the Development of High Fat Diet Induced Non-Alcoholic Steatohepatitis. *Toxicology and Applied Pharmacology*, **330**, 1-8. <https://doi.org/10.1016/j.taap.2017.06.023>
- [29] Miyata, M., Sakaida, Y., Matsuzawa, H., *et al.* (2011) Fibroblast Growth Factor 19 Treatment Ameliorates Disruption of Hepatic Lipid Metabolism in Farnesoid X Receptor (Fxr)-Null Mice. *Biological and Pharmaceutical Bulletin*, **34**, 1885-1889.
- [30] Alvarez-Sola, G., Uriarte, I., Latasa, M.U., *et al.* (2017) Fibroblast Growth Factor 15/19 (FGF15/19) Protects from Diet-Induced Hepatic Steatosis: Development of an FGF19-Based Chimeric Molecule to Promote Fatty Liver Regeneration. *Gut*, **66**, 1818-1828. <https://doi.org/10.1136/gutjnl-2016-312975>
- [31] Zhou, M., Learned, R.M., Rossi, S.J., *et al.* (2017) Engineered FGF19 Eliminates Bile Acid Toxicity and Lipotoxicity Leading to Resolution of Steatohepatitis and Fibrosis in Mice. *Hepatology Communications*, **1**, 1024-1042. <https://doi.org/10.1002/hep4.1108>
- [32] Uriarte, I., Latasa, M.U., Carotti, S., *et al.* (2015) Ileal FGF15 Contributes to Fibrosis-Associated Hepatocellular Carcinoma Development. *International Journal of Cancer*, **136**, 2469-2475. <https://doi.org/10.1002/ijc.29287>
- [33] Harrison, S.A., Rinella, M.E., Abdelmalek, M.F., *et al.* (2018) NGM282 for Treatment of Non-Alcoholic Steatohepatitis: A Multicentre, Randomised, Double-Blind, Placebo-Controlled, Phase 2 Trial. *The Lancet*, **391**, 1174-1185. [https://doi.org/10.1016/S0140-6736\(18\)30474-4](https://doi.org/10.1016/S0140-6736(18)30474-4)
- [34] Harrison, S., Rossi, S., Bashir, M., *et al.* (2018) NGM282 Improves Fibrosis and NASH-Related Histology in 12 Weeks in Patients with Biopsy-Confirmed NASH, Which Is Preceded by Significant Decreases in Hepatic Steatosis, Liver Transaminases and Fibrosis Markers at 6 Weeks. *Journal of Hepatology*, **68**, S65-S66. [https://doi.org/10.1016/S0168-8278\(18\)30352-0](https://doi.org/10.1016/S0168-8278(18)30352-0)
- [35] Harrison, S.A., Neff, G., Guy, C.D., *et al.* (2021) Efficacy and Safety of Aldafermin, an Engineered FGF19 Analog, in a Randomized, Double-Blind, Placebo-Controlled Trial of Patients with Nonalcoholic Steatohepatitis. *Gastroenterology*, **160**, 219-231.E1. <https://doi.org/10.1053/j.gastro.2020.08.004>
- [36] Lan, T., Morgan, D.A., Rahmouni, K., *et al.* (2017) FGF19, FGF21, and an FGFR1/ $\beta$ -Klotho-Activating Antibody Act on the Nervous System to Regulate Body Weight and Glycemia. *Cell Metabolism*, **26**, 709-718.E3. <https://doi.org/10.1016/j.cmet.2017.09.005>
- [37] Wu, A.L., Coulter, S., Liddle, C., *et al.* (2011) FGF19 Regulates Cell Proliferation, Glucose and Bile Acid Metabolism via FGFR4-Dependent and Independent Pathways. *PLoS ONE*, **6**, e17868. <https://doi.org/10.1371/journal.pone.0017868>
- [38] Sun, Y.N., Yang, Z.X., Ren, F.Z. and Fang, B. (2020) FGF19 Alleviates Palmitate-Induced Atrophy in C2C12 Cells by Inhibiting Mitochondrial Overload and Insulin Resistance. *International Journal of Biological Macromolecules*, **158**, 401-407. <https://doi.org/10.1016/j.ijbiomac.2020.04.186>
- [39] Guo, A., Li, K. and Xiao, Q. (2020) Fibroblast Growth Factor 19 Alleviates Palmitic Acid-Induced Mitochondrial Dysfunction and Oxidative Stress via the AMPK/PGC-1 $\alpha$  Pathway in Skeletal Muscle. *Biochemical and Biophysical Research Communications*, **526**, 1069-1076. <https://doi.org/10.1016/j.bbrc.2020.04.002>
- [40] Kong, B., Sun, R., Huang, M., *et al.* (2018) Fibroblast Growth Factor 15-Dependent and Bile Acid-Independent Promotion of Liver Regeneration in Mice. *Hepatology*, **68**, 1961-1976. <https://doi.org/10.1002/hep.30041>
- [41] French, D.M., Lin, B.C., Wang, M.P., *et al.* (2012) Targeting FGFR4 Inhibits Hepatocellular Carcinoma in Preclinical Mouse Models. *PLoS ONE*, **7**, e36713. <https://doi.org/10.1371/journal.pone.0036713>
- [42] Tan, Q., Li, F., Wang, G., *et al.* (2016) Identification of FGF19 as a Prognostic Marker and Potential Driver Gene of Lung Squamous Cell Carcinomas in Chinese Smoking Patients. *Oncotarget*, **7**, 18394-402. <https://doi.org/10.18632/oncotarget.7817>
- [43] Li, F., Li, Z., Han, Q., *et al.* (2020) Enhanced Autocrine FGF19/FGFR4 Signaling Drives the Progression of Lung Squamous Cell Carcinoma, Which Responds to mTOR Inhibitor AZD2104. *Oncogene*, **39**, 3507-3521. <https://doi.org/10.1038/s41388-020-1227-2>
- [44] Wu, X., Ge, H., Lemon, B., *et al.* (2010) Separating Mitogenic and Metabolic Activities of Fibroblast Growth Factor

- 
- 19 (FGF19). *Proceedings of the National Academy of Sciences of the United States of America*, **107**, 14158-14163. <https://doi.org/10.1073/pnas.1009427107>
- [45] Benoit, B., Meugnier, E., Castelli, M., *et al.* (2017) Fibroblast Growth Factor 19 Regulates Skeletal Muscle Mass and Ameliorates Muscle Wasting in Mice. *Nature Medicine*, **23**, 990-996. <https://doi.org/10.1038/nm.4363>
- [46] Guo, A., Li, K., Tian, H.C., *et al.* (2021) FGF19 Protects Skeletal Muscle against Obesity-Induced Muscle Atrophy, Metabolic Derangement and Abnormal Irisin Levels via the AMPK/SIRT-1/PGC- $\alpha$  Pathway. *Journal of Cellular and Molecular Medicine*, **25**, 3585-3600. <https://doi.org/10.1111/jcmm.16448>
- [47] Itoh, N., Ohta, H., Nakayama, Y. and Konishi, M. (2016) Roles of FGF Signals in Heart Development, Health, and Disease. *Frontiers in Cell and Developmental Biology*, **4**, Article No. 110. <https://doi.org/10.3389/fcell.2016.00110>
- [48] Maeda, T., Kanzaki, H., Chiba, T., *et al.* (2019) Serum Fibroblast Growth Factor 19 Serves as a Potential Novel Biomarker for Hepatocellular Carcinoma. *BMC Cancer*, **19**, Article No. 1088. <https://doi.org/10.1186/s12885-019-6322-9>