

非小细胞肺癌免疫微环境的CT与PET/CT组学研究进展

黄晓舟¹, 钱丹飞¹, 杨建峰^{2*}

¹绍兴文理学院医学院, 浙江 绍兴

²绍兴市人民医院(绍兴文理学院附属第一医院)放射科, 浙江 绍兴

收稿日期: 2025年7月12日; 录用日期: 2025年8月5日; 发布日期: 2025年8月14日

摘要

免疫疗法已在非小细胞肺癌(NSCLC)治疗中发挥重要作用。肿瘤免疫微环境特征强烈影响NSCLC免疫治疗的潜在反应, 准确评估NSCLC免疫微环境状态对制定免疫治疗策略和评估患者预后具有重要意义。CT影像组学利用定量分析纹理特征反映肿瘤异质性, 已被应用于评估免疫微环境的研究。本文就影像组学在NSCLC免疫微环境研究中的应用现状进行综述, 并对其应用前景进行展望。

关键词

非小细胞肺癌, 免疫微环境, 影像组学, 肿瘤浸润淋巴细胞

Research Progress in CT and PET/CT Radiomics of the Tumor Immune Microenvironment in Non-Small Cell Lung Cancer

Xiaozhou Huang¹, Danfei Qian¹, Jianfeng Yang^{2*}

¹School of Medicine, Shaoxing University, Shaoxing Zhejiang

²Department of Radiology, Shaoxing People's Hospital (The First Affiliated Hospital of Shaoxing University), Shaoxing Zhejiang

Received: Jul. 12th, 2025; accepted: Aug. 5th, 2025; published: Aug. 14th, 2025

*通讯作者。

文章引用: 黄晓舟, 钱丹飞, 杨建峰. 非小细胞肺癌免疫微环境的CT与PET/CT组学研究进展[J]. 临床医学进展, 2025, 15(8): 942-949. DOI: 10.12677/acm.2025.1582319

Abstract

Immunotherapy has played a pivotal role in the treatment of non-small cell lung cancer (NSCLC). The characteristics of the tumor immune microenvironment (TIME) significantly influence potential responses to immunotherapy in NSCLC, making accurate assessment of TIME status crucial for developing treatment strategies and evaluating patient prognosis. CT radiomics, through quantitative analysis of texture features that reflect tumor heterogeneity, has been increasingly applied in TIME evaluation research. This review summarizes current applications of radiomics in NSCLC TIME studies and discusses future perspectives.

Keywords

Non-Small Cell Lung Cancer, Tumor Immune Microenvironment, Radiomics, Tumor-Infiltrating Lymphocytes

Copyright © 2025 by author(s) and Hans Publishers Inc.

This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

1. 介绍

非小细胞肺癌(NSCLC)是最常见的肺癌类型, 5年生存率仅为15% [1]。随着NSCLC免疫治疗的临床应用, 肿瘤微环境(TME)研究越来越受到重视。肿瘤微环境(Tumor microenvironment, TME)包括网状纤维母细胞、血管内皮细胞、淋巴管上皮细胞、细胞外基质等间质细胞建立的微环境结构和不同亚型的T、B淋巴细胞、树突状细胞等免疫细胞浸润的免疫微环境[2], 在肿瘤发生、生长、侵袭、转移和治疗反应方面发挥重要影响[3] [4]。研究表明肿瘤免疫微环境(TIME)表型(例如被CD8⁺ T细胞浸润的肿瘤)与免疫治疗的疗效和生存率呈正相关[5]-[8]。评估TIME各种成分的相互作用和免疫逃逸机制是进一步改善当前免疫疗法或开发新治疗方法的关键[9]。通过活检或手术取得病理标本是检测确定TIME表型的金标准, 但存在肿瘤细胞播撒、术中出血、采样误差, 可重复性差无法动态评估等缺陷[10]-[14]。

影像学组学从CT、MR、超声等图像中提取、整合特征, 诊断并预测肿瘤组织学亚型、基因突变状态、基因表达水平以及临床预后[15]。CT通过定量形态学及纹理特征揭示肿瘤异质性, 间接反映微环境物理特性(如坏死、纤维化)及免疫浸润状态, 但其功能信息局限, 无法直接捕捉肿瘤的代谢活性或免疫细胞的动态分布。PET/CT结合了CT的解剖信息和PET的功能代谢信息, 尤其通过18F-FDG代谢参数(如SUV_{max}、MTV)直接评估肿瘤葡萄糖代谢活性。研究表明, 高SUV_{max}可能与“热”肿瘤(即高免疫细胞浸润)相关, 而低SUV_{max}可能提示“冷”肿瘤(免疫排斥表型)。此外, PET/CT可通过动态扫描评估肿瘤代谢的动态变化, 为免疫治疗响应提供早期预测。但也存在辐射高、成本昂贵及免疫亚群特异性不足的缺点。CT的纹理特征(如GLNU)可量化肿瘤的结构异质性, 而PET的代谢参数可反映肿瘤的生物学活性。两者结合可更全面地描绘肿瘤的“冷”“热”表型。PET/CT的动态扫描可捕捉治疗前后代谢变化, 而CT的稳定性特征可提供基线结构信息, 两者结合有助于区分治疗引起的免疫激活与假性进展。

目前, 对NSCLC的组学特征与TIME之间的相关性已得到证实。基于机器学习等放射组学模型已经得到广泛应用, 影像学评估TIME在临床实践中发挥重要的作用。本文就CT组学在NSCLC的TIME应用现状进行综述, 并探讨其应用前景。

2. 程序性死亡 1/程序性死亡配体 1

在肿瘤免疫过程中,程序性死亡 1 (PD-1)/程序性死亡配体 1 (PD-L1)通路调节肿瘤浸润淋巴细胞(TIL)的活化。由于 T 细胞表面的 PD-1 受体与其配体(PD-L1)结合阻断 T 细胞活化及随后的免疫应答,使肿瘤细胞逃避免疫系统的攻击,导致预后较差[16]-[18]。因此,通过抗 PD-1 或抗 PD-L1 药物抑制 PD-1 和 PD-L1 结合恢复 T 细胞的免疫功能,从而提供良好的抗肿瘤效果[19] [20]。近年来的研究表明,抗 PD-1/PD-L1 单克隆抗体形式的免疫检查点抑制剂(ICI)已成为治疗 NSCLC 的基石,且在晚期转移性 NSCLC 患者的治疗中也显示出前景[21]-[23]。

PD-L1 的表达与 NSCLC 患者的预后相关,研究表明 PD-L1 表达阴性(肿瘤比例评分(TPS) < 1%)的患者不适合抗 PD-L1 抗体治疗,而 PD-L1 表达阳性(TPS \geq 1%)的患者则可以从抗 PD-L1 抗体中获益[24] [25]。影像组学特征(如纹理特征、灰度共生矩阵特征)能够反映肿瘤内部的异质性,这种异质性可能与肿瘤微环境的病理生理学特征密切相关。例如,高熵值(Entropy)可能提示肿瘤内部坏死或纤维化区域的增加,而低对比度(Contrast)可能反映细胞密集区域的均质性[26] [27]。这些微环境特征可能通过影响免疫细胞的分布和功能,进一步调控 PD-L1 的表达。研究表明,肿瘤内部的坏死区域可能通过释放损伤相关分子模式(DAMPs)激活免疫应答,而纤维化区域则可能通过物理屏障限制 T 细胞浸润[28]。目前,NSCLC 患者 PD-L1 的放射组学研究主要集中于评估 PD-L1 表达和抗 PD-1/PD-L1 治疗反应的预测[29]-[38]。Weng 等研究者基于 CT 影像组学、临床病理学以及 CT 形态学特征参数建立放射组学模型,预测 120 例晚期 NSCLC 患者的 PD-L1 表达水平和肿瘤突变负荷(TMB)状态的受试者工作特征曲线下面积(AUC)分别为 0.839 和 0.818 [29]。Mu 等学者分析两家机构 210 例接受免疫检查点抑制剂(ICI)治疗的 NSCLC 患者 PET/CT 图像的放射组学特征和临床数据建立放射组学模型预测 NSCLC 患者患恶液质 AUC \geq 0.74 [30]。此外,Li 等从 PET/CT 图像中提取影像组学特征和深度学习特征建立基于所选特征的融合模型,预测 136 例 NSCLC 患者的 PD-L1 表达,在训练和验证队列中 AUC 分别为 0.954 和 0.910 [31]。上述研究表明,影像组学预测模型可以个性化预测 NSCLC 患者 PD-L1 表达,筛选抗 PD-L1 免疫治疗获益的患者。

3. 肿瘤浸润淋巴细胞

肿瘤浸润淋巴细胞(TIL),特别是 CD8⁺ TIL 及其免疫调节细胞因子代表适应性免疫,在 NSCLC 免疫微环境中执行关键效应细胞毒性功能并介导 ICI 应答[39]。既往研究表明,通过显示 CD8⁺ TIL 是否高浸润可评估抗肿瘤反应,并与 ICI 的临床应答相关,可作为判断患者预后的生物标志物[40]。由于 NSCLC 是一种高度时空异质性的疾病,且存在病理学采样偏差,在标本中评估的 TIL 可能无法代表整个肿瘤中的 TIL 水平[41]。

影像组学特征对肿瘤浸润淋巴细胞(TIL)的预测效能可能与其对肿瘤微环境物理特征的量化能力有关。例如,CT 图像中的高灰度值不均匀性(Gray-Level Non-Uniformity, GLNU)可能反映肿瘤内部的血管密度差异,而血管密度是影响 T 细胞浸润的关键因素之一[42]。此外,肿瘤内部的低密度区域(如囊变或坏死)可能通过减少 T 细胞的趋化和存活,降低 CD8⁺ T 细胞的浸润水平[43]。目前,影像组学研究主要集中于 CD8⁺ T 细胞并取得较满意的结果[38] [44]-[48]。Tong 等[44]分析 221 例 NSCLC 患者的 18F-FDG PET/CT 图像,并建立影像组学标记和临床特征联合预测模型,结果显示该模型在训练组和验证组中的 AUC 分别为 0.93 和 0.92,有效预测 NSCLC 患者 CD8 的表达。Chen 等[45]从 117 例 NSCLC 患者的 CT 图像中提取强 CD8⁺ TIL 丰度的影像组学特征构建影像组学评分,研究证明基于 CT 的影像组学特征对 NSCLC 患者原发肿瘤病灶的 CD8⁺ TIL 具有良好的预测效能,有望成为免疫治疗患者潜在生物标志物。此外,Mazzaschi 等[46] [47]建立基于 TIME 图谱的影像组学模型预测 NSCLC 患者的预后,并为晚期 NSCLC 病例提供可利用的预测策略。

近年的研究表明高 CD3 T 细胞计数是对 PD-1 阻断反应的独立预测因子[49]。总体 CD3 T 细胞计数较高与无进展生存期长具有相关性[50]。CD3 T 细胞具有抗肿瘤活性,对总生存期和复发率具有极大的预后意义,并且是重要的预后预测指标[51]-[53]。Chen 等[54]研究者基于 CT 组学特征和患者的临床病理信息建立放射组学模型,预测 105 例 NSCLC 患者的 CD3 T 细胞的表达的 AUC 为 0.94,在验证集中 AUC 为 0.73。

目前,对 NSCLC 中肿瘤浸润淋巴细胞的放射组学研究正持续推进,且大多数模型已取得良好的预测性能。研究主要包括两个方面:术前放射组学无创性预测 TIL 表达和放射组学特征结合 TIL 预测预后。总体上,NSCLC 肿瘤浸润淋巴细胞的放射组学研究已经启动,但是大多数研究集中在 CD8⁺ T 细胞,需要对其他类型的 TIL 进行更多的研究。另外,TIL 研究以单中心以探索性、概念验证的方式进行,需要整合来自多个中心的数据提高这些预测模型临床可行性应用。

4. 影像组学在其他免疫微环境成分研究中的应用

影像组学还用于对自然杀伤(NK)细胞、接受放射治疗后的中性粒细胞与淋巴细胞比率(NLR)的预测进行建模。研究表明,抑制 IL6 可增强奥斯替尼耐药 EGFR 突变型 NSCLC 细胞中 NK 细胞介导的细胞毒性[55]。另一项研究表明,NSCLC 中 SERPINB 4 的上调抑制 NK 细胞介导的细胞毒性,是调节 NSCLC 的免疫应答的潜在治疗途径[56]。这些发现强调 NK 细胞在 NSCLC 中协调抗肿瘤反应的关键作用,并凸显以 NK 细胞为中心的治疗策略的新途径。Meng 等人开发了一种基于增强 CT 图像的放射组学模型,能够准确预测 NK 细胞在 NSCLC 中的浸润情况,具有良好的稳定性和诊断效能[57]。Hou 等人开发了一种基于三维(3D)剂量分布图、CT 图像以及临床特征的剂量组学&影像组学&临床融合模型,预测了 242 例局部晚期非小细胞肺癌(LA-NSCLC)患者放疗后 NLR,其 AUC 为 0.765。该模型可为放射治疗相关炎症反应的评价提供参考,对指导治疗方案的优化具有一定的应用价值[58]。

以上研究表明,影像组学在评估 NK 细胞、NLR 表达方面具有良好预测结果。证明影像组学具有评价其他 TIME 组分的潜力。在未来的研究中,同样需要扩大研究范围和样本量,以获得更准确和全面的 TIME 预测模型。

5. 问题和挑战

影像组学为 NSCLC 患者 TIME 的非侵入性评估提供了新思路,有助于治疗决策和预后判断,但其临床应用仍面临诸多挑战。

首先,影像组学模型建立需要预先在图像上手动分割病灶,复杂、耗时且主观,可靠性及可重复性难以保证,限制了其在临床上的推广。未来通过对尝试自动分割的研究可能会提高临床适用性,提高分割速度和准确性[59]。

再者,再现性仍然是现有影像组学模型的主要障碍[60]。对给定扫描仪内的稳定体模对象进行的重新测试研究估计,仅约 30%的 MRI 特征具有再现性,而多扫描仪体模研究显示特征再现性范围为 15%~85% [61]。多数影像组学特征缺乏明确的生物学解释,或仅反映技术差异及混杂因素。因此,有必要建立一致的方案,以避免数据采集的差异性。使用强度标准化,对感兴趣区进行异常值剔除,并采用固定的 bin 大小的灰度离散化,可以提高特征的可重复性[62]。

许多影像组学模型可能仅仅是基于训练数据集大小和可用特征数量之间的差异而过拟合其训练数据。因此,模型未能很好地推广到其他数据集。目前,基于影像组学对 NSCLC 的 TIME 评价多为单中心、小样本研究,结果尚未得到外部验证,这可能会对模型适应,优化和评估的过程产生负面影响[63]。因此,未来需开展多中心、标准化的大规模前瞻性研究,以系统性评估影像组学在 NSCLC 肿瘤微环境分析中的

实际效能。

尽管影像组学在预测免疫微环境状态方面显示出潜力，但其生物学解释仍需进一步明确。例如，某些纹理特征可能同时反映多种病理生理学过程(如坏死、纤维化或血管增生)，导致模型的可解释性降低。未来的研究应结合多组学数据，以明确影像组学特征与特定微环境特征(如免疫细胞空间分布或细胞外基质组成)的关联，从而提升模型的生物学合理性和临床适用性。

6. 结论

总之，精确成像已成为医学影像学的常态，影像组学的发展为非小细胞肺癌 TIME 的无创评价提供新的思路，同时也开创个性化 TIME 评价的新领域。目前，研究表明影像组学具有评估(上述的 PD-1/PD-L1, CD8⁺, CD3 TIL, NK 细胞、NLR) TIME 的潜力，并且可以预期未来研究的突破。通过建立大型数据库和开展多中心合作，可以进一步推动影像组学的研究，更好地分析 TIME，并最终成为临床认可的无创评估方法。近年来，随着人工智能技术的快速发展，尤其是机器学习和深度学习领域的突破性进展，肺部疾病的影像诊断技术取得了显著进步。未来研究应整合影像组学与遗传学、分子生物学及病理学等多学科数据，以提升对恶性肿瘤预后评估的准确性。这种多学科融合的方法将为临床治疗方案的选择和优化提供科学依据，最终推动个体化精准医疗的实现。

基金项目

浙江省医药卫生科技计划一般项目(2023KY1236); 绍兴市科技计划基础公益类项目(2022A14010)。

参考文献

- [1] Goldstraw, P., Ball, D., Jett, J.R., Le Chevalier, T., Lim, E., Nicholson, A.G., *et al.* (2011) Non-Small-Cell Lung Cancer. *The Lancet*, **378**, 1727-1740. [https://doi.org/10.1016/s0140-6736\(10\)62101-0](https://doi.org/10.1016/s0140-6736(10)62101-0)
- [2] Shintani, Y., Kimura, T., Funaki, S., Ose, N., Kanou, T. and Fukui, E. (2023) Therapeutic Targeting of Cancer-Associated Fibroblasts in the Non-Small Cell Lung Cancer Tumor Microenvironment. *Cancers*, **15**, Article 335. <https://doi.org/10.3390/cancers15020335>
- [3] Pellat, A. and Barat, M. (2023) Tumor Microenvironment: A New Application for Radiomics. *Diagnostic and Interventional Imaging*, **104**, 93-94. <https://doi.org/10.1016/j.diii.2022.10.011>
- [4] Jin, M. and Jin, W. (2020) The Updated Landscape of Tumor Microenvironment and Drug Repurposing. *Signal Transduction and Targeted Therapy*, **5**, Article No. 166. <https://doi.org/10.1038/s41392-020-00280-x>
- [5] Binnewies, M., Roberts, E.W., Kersten, K., Chan, V., Fearon, D.F., Merad, M., *et al.* (2018) Understanding the Tumor Immune Microenvironment (TIME) for Effective Therapy. *Nature Medicine*, **24**, 541-550. <https://doi.org/10.1038/s41591-018-0014-x>
- [6] Chen, D.S. and Mellman, I. (2017) Elements of Cancer Immunity and the Cancer-Immune Set Point. *Nature*, **541**, 321-330. <https://doi.org/10.1038/nature21349>
- [7] Gajewski, T.F., Corrales, L., Williams, J., Horton, B., Sivan, A. and Spranger, S. (2017) Cancer Immunotherapy Targets Based on Understanding the T Cell-Inflamed versus Non-T Cell-Inflamed Tumor Microenvironment. In: Kalinski, P., Ed., *Tumor Immune Microenvironment in Cancer Progression and Cancer Therapy*, Springer, 19-31. https://doi.org/10.1007/978-3-319-67577-0_2
- [8] Galon, J. and Bruni, D. (2019) Approaches to Treat Immune Hot, Altered and Cold Tumours with Combination Immunotherapies. *Nature Reviews Drug Discovery*, **18**, 197-218. <https://doi.org/10.1038/s41573-018-0007-y>
- [9] Ugel, S., Canè, S., De Sanctis, F. and Bronte, V. (2021) Monocytes in the Tumor Microenvironment. *Annual Review of Pathology: Mechanisms of Disease*, **16**, 93-122. <https://doi.org/10.1146/annurev-pathmechdis-012418-013058>
- [10] McGranahan, N. and Swanton, C. (2017) Clonal Heterogeneity and Tumor Evolution: Past, Present, and the Future. *Cell*, **168**, 613-628. <https://doi.org/10.1016/j.cell.2017.01.018>
- [11] McLaughlin, J., Han, G., Schalper, K.A., Carvajal-Hausdorf, D., Pelekanou, V., Rehman, J., *et al.* (2016) Quantitative Assessment of the Heterogeneity of PD-L1 Expression in Non-Small-Cell Lung Cancer. *JAMA Oncology*, **2**, 46-54. <https://doi.org/10.1001/jamaoncol.2015.3638>

- [12] Jiménez-Sánchez, A., Memon, D., Pourpe, S., Veeraraghavan, H., Li, Y., Vargas, H.A., *et al.* (2017) Heterogeneous Tumor-Immune Microenvironments among Differentially Growing Metastases in an Ovarian Cancer Patient. *Cell*, **170**, 927-938.e20. <https://doi.org/10.1016/j.cell.2017.07.025>
- [13] Mansfield, A.S., Aubry, M.C., Moser, J.C., Harrington, S.M., Dronca, R.S., Park, S.S., *et al.* (2016) Temporal and Spatial Discordance of Programmed Cell Death-Ligand 1 Expression and Lymphocyte Tumor Infiltration between Paired Primary Lesions and Brain Metastases in Lung Cancer. *Annals of Oncology*, **27**, 1953-1958. <https://doi.org/10.1093/annonc/mdw289>
- [14] Santos, R., Ursu, O., Gaulton, A., Bento, A.P., Donadi, R.S., Bologa, C.G., *et al.* (2016) A Comprehensive Map of Molecular Drug Targets. *Nature Reviews Drug Discovery*, **16**, 19-34. <https://doi.org/10.1038/nrd.2016.230>
- [15] Chen, P., Liu, Y., Wen, Y. and Zhou, C. (2022) Non-Small Cell Lung Cancer in China. *Cancer Communications*, **42**, 937-970. <https://doi.org/10.1002/cac2.12359>
- [16] Incorvaia, L., Fanale, D., Badalamenti, G., Barraco, N., Bono, M., Corsini, L.R., *et al.* (2019) Programmed Death Ligand 1 (PD-L1) as a Predictive Biomarker for Pembrolizumab Therapy in Patients with Advanced Non-Small-Cell Lung Cancer (NSCLC). *Advances in Therapy*, **36**, 2600-2617. <https://doi.org/10.1007/s12325-019-01057-7>
- [17] Chen, L. and Han, X. (2015) Anti-PD-1/PD-L1 Therapy of Human Cancer: Past, Present, and Future. *Journal of Clinical Investigation*, **125**, 3384-3391. <https://doi.org/10.1172/jci80011>
- [18] Riley, J.L. (2009) PD-1 Signaling in Primary T Cells. *Immunological Reviews*, **229**, 114-125. <https://doi.org/10.1111/j.1600-065x.2009.00767.x>
- [19] Lages, C.S., Lewkowich, I., Sproles, A., Wills-Karp, M. and Chougnet, C. (2010) Partial Restoration of T-Cell Function in Aged Mice by *in Vitro* Blockade of the PD-1/PD-L1 Pathway. *Aging Cell*, **9**, 785-798. <https://doi.org/10.1111/j.1474-9726.2010.00611.x>
- [20] Taube, J.M., Klein, A., Brahmer, J.R., Xu, H., Pan, X., Kim, J.H., *et al.* (2014) Association of PD-1, PD-1 Ligands, and Other Features of the Tumor Immune Microenvironment with Response to Anti-PD-1 Therapy. *Clinical Cancer Research*, **20**, 5064-5074. <https://doi.org/10.1158/1078-0432.ccr-13-3271>
- [21] 朱闻捷, 朱豪华, 刘雨桃, 等. 程序性死亡蛋白 1/程序性死亡蛋白配体 1 抑制剂治疗晚期非小细胞肺癌的疗效及疗效和预后预测标志物的真实世界研究[J]. 中华肿瘤杂志, 2022, 44(5): 416-424.
- [22] Hanna, N., Johnson, D., Temin, S., Baker, S., Brahmer, J., Ellis, P.M., *et al.* (2017) Systemic Therapy for Stage IV Non-Small-Cell Lung Cancer: American Society of Clinical Oncology Clinical Practice Guideline Update. *Journal of Clinical Oncology*, **35**, 3484-3515. <https://doi.org/10.1200/jco.2017.74.6065>
- [23] Rocco, D., Malapelle, U., Del Re, M., Della Gravara, L., Pepe, F., Danesi, R., *et al.* (2020) Pharmacodynamics of Current and Emerging PD-1 and PD-L1 Inhibitors for the Treatment of Non-Small Cell Lung Cancer. *Expert Opinion on Drug Metabolism & Toxicology*, **16**, 87-96. <https://doi.org/10.1080/17425255.2020.1721460>
- [24] Doroshow, D.B., Bhalla, S., Beasley, M.B., Sholl, L.M., Kerr, K.M., Gnjatic, S., *et al.* (2021) PD-L1 as a Biomarker of Response to Immune-Checkpoint Inhibitors. *Nature Reviews Clinical Oncology*, **18**, 345-362. <https://doi.org/10.1038/s41571-021-00473-5>
- [25] Mok, T.S.K., Wu, Y., Kudaba, I., Kowalski, D.M., Cho, B.C., Turna, H.Z., *et al.* (2019) Pembrolizumab versus Chemotherapy for Previously Untreated, Pd-L1-Expressing, Locally Advanced or Metastatic Non-Small-Cell Lung Cancer (KEYNOTE-042): A Randomised, Open-Label, Controlled, Phase 3 Trial. *The Lancet*, **393**, 1819-1830. [https://doi.org/10.1016/s0140-6736\(18\)32409-7](https://doi.org/10.1016/s0140-6736(18)32409-7)
- [26] Lambin, P., Rios-Velazquez, E., Leijenaar, R., Carvalho, S., van Stiphout, R.G.P.M., Granton, P., *et al.* (2012) Radiomics: Extracting More Information from Medical Images Using Advanced Feature Analysis. *European Journal of Cancer*, **48**, 441-446. <https://doi.org/10.1016/j.ejca.2011.11.036>
- [27] Aerts, H.J.W.L., Velazquez, E.R., Leijenaar, R.T.H., Parmar, C., Grossmann, P., Carvalho, S., *et al.* (2014) Decoding Tumor Phenotype by Noninvasive Imaging Using a Quantitative Radiomics Approach. *Nature Communications*, **5**, Article No. 4006. <https://doi.org/10.1038/ncomms5006>
- [28] Liu, Z., Wang, S., Dong, D., Wei, J., Fang, C., Zhou, X., *et al.* (2019) The Applications of Radiomics in Precision Diagnosis and Treatment of Oncology: Opportunities and Challenges. *Theranostics*, **9**, 1303-1322. <https://doi.org/10.7150/thno.30309>
- [29] Wen, Q., Yang, Z., Dai, H., Feng, A. and Li, Q. (2021) Radiomics Study for Predicting the Expression of PD-L1 and Tumor Mutation Burden in Non-Small Cell Lung Cancer Based on CT Images and Clinicopathological Features. *Frontiers in Oncology*, **11**, Article 620246. <https://doi.org/10.3389/fonc.2021.620246>
- [30] Mu, W., Katsoulakis, E., Whelan, C.J., Gage, K.L., Schabath, M.B. and Gillies, R.J. (2021) Radiomics Predicts Risk of Cachexia in Advanced NSCLC Patients Treated with Immune Checkpoint Inhibitors. *British Journal of Cancer*, **125**, 229-239. <https://doi.org/10.1038/s41416-021-01375-0>

- [31] Li, B., Su, J., Liu, K. and Hu, C. (2024) Deep Learning Radiomics Model Based on PET/CT Predicts PD-L1 Expression in Non-Small Cell Lung Cancer. *European Journal of Radiology Open*, **12**, Article ID: 100549. <https://doi.org/10.1016/j.ejro.2024.100549>
- [32] Jiang, M., Sun, D., Guo, Y., Guo, Y., Xiao, J., Wang, L., *et al.* (2020) Assessing PD-L1 Expression Level by Radiomic Features from PET/CT in Nonsmall Cell Lung Cancer Patients: An Initial Result. *Academic Radiology*, **27**, 171-179. <https://doi.org/10.1016/j.acra.2019.04.016>
- [33] Vaidya, P., Bera, K., Patil, P.D., Gupta, A., Jain, P., Alilou, M., *et al.* (2020) Novel, Non-Invasive Imaging Approach to Identify Patients with Advanced Non-Small Cell Lung Cancer at Risk of Hyperprogressive Disease with Immune Checkpoint Blockade. *Journal for ImmunoTherapy of Cancer*, **8**, e001343. <https://doi.org/10.1136/jitc-2020-001343>
- [34] Liu, Y., Wu, M., Zhang, Y., Luo, Y., He, S., Wang, Y., *et al.* (2021) Imaging Biomarkers to Predict and Evaluate the Effectiveness of Immunotherapy in Advanced Non-Small-Cell Lung Cancer. *Frontiers in Oncology*, **11**, Article 657615. <https://doi.org/10.3389/fonc.2021.657615>
- [35] Bracci, S., Dolciemi, M., Trobiani, C., Izzo, A., Pernazza, A., D'Amati, G., *et al.* (2021) Quantitative CT Texture Analysis in Predicting PD-L1 Expression in Locally Advanced or Metastatic NSCLC Patients. *La radiologia medica*, **126**, 1425-1433. <https://doi.org/10.1007/s11547-021-01399-9>
- [36] Ren, Q., Xiong, F., Zhu, P., Chang, X., Wang, G., He, N., *et al.* (2022) Assessing the Robustness of Radiomics/Deep Learning Approach in the Identification of Efficacy of Anti-PD-1 Treatment in Advanced or Metastatic Non-Small Cell Lung Carcinoma Patients. *Frontiers in Oncology*, **12**, Article 952749. <https://doi.org/10.3389/fonc.2022.952749>
- [37] Wang, C., Ma, J., Shao, J., Zhang, S., Liu, Z., Yu, Y., *et al.* (2022) Predicting EGFR and PD-L1 Status in NSCLC Patients Using Multitask AI System Based on CT Images. *Frontiers in Immunology*, **13**, Article 813072. <https://doi.org/10.3389/fimmu.2022.813072>
- [38] Zhou, J., Zou, S., Kuang, D., Yan, J., Zhao, J. and Zhu, X. (2021) A Novel Approach Using FDG-PET/CT-Based Radiomics to Assess Tumor Immune Phenotypes in Patients with Non-Small Cell Lung Cancer. *Frontiers in Oncology*, **11**, Article 769272. <https://doi.org/10.3389/fonc.2021.769272>
- [39] Altorki, N.K., Markowitz, G.J., Gao, D., Port, J.L., Saxena, A., Stiles, B., *et al.* (2018) The Lung Microenvironment: An Important Regulator of Tumour Growth and Metastasis. *Nature Reviews Cancer*, **19**, 9-31. <https://doi.org/10.1038/s41568-018-0081-9>
- [40] Tumei, P.C., Harview, C.L., Yearley, J.H., Shintaku, I.P., Taylor, E.J.M., Robert, L., *et al.* (2014) PD-1 Blockade Induces Responses by Inhibiting Adaptive Immune Resistance. *Nature*, **515**, 568-571. <https://doi.org/10.1038/nature13954>
- [41] Tozaki, M., Sakamoto, M., Oyama, Y., Maruyama, K. and Fukuma, E. (2010) Predicting Pathological Response to Neoadjuvant Chemotherapy in Breast Cancer with Quantitative ¹H MR Spectroscopy Using the External Standard Method. *Journal of Magnetic Resonance Imaging*, **31**, 895-902. <https://doi.org/10.1002/jmri.22118>
- [42] Xie, W., Jiang, S., Xin, F., Jiang, Z., Pan, W., Zhou, X., *et al.* (2024) Prediction of CD8⁺ T Lymphocyte Infiltration Levels in Gastric Cancer from Contrast-Enhanced CT and Clinical Factors Using Machine Learning. *Medical Physics*, **51**, 7108-7118. <https://doi.org/10.1002/mp.17350>
- [43] Liao, H., Zhang, Z., Chen, J., Liao, M., Xu, L., Wu, Z., *et al.* (2019) Preoperative Radiomic Approach to Evaluate Tumor-Infiltrating CD8⁺ T Cells in Hepatocellular Carcinoma Patients Using Contrast-Enhanced Computed Tomography. *Annals of Surgical Oncology*, **26**, 4537-4547. <https://doi.org/10.1245/s10434-019-07815-9>
- [44] Tong, H., Sun, J., Fang, J., Zhang, M., Liu, H., Xia, R., *et al.* (2022) A Machine Learning Model Based on PET/CT Radiomics and Clinical Characteristics Predicts Tumor Immune Profiles in Non-Small Cell Lung Cancer: A Retrospective Multicohort Study. *Frontiers in Immunology*, **13**, Article 859323. <https://doi.org/10.3389/fimmu.2022.859323>
- [45] Chen, Y., Xu, T., Jiang, C., You, S., Cheng, Z. and Gong, J. (2022) CT-Based Radiomics Signature to Predict CD8⁺ Tumor Infiltrating Lymphocytes in Non-Small-Cell Lung Cancer. *Acta Radiologica*, **64**, 1390-1399. <https://doi.org/10.1177/02841851221126596>
- [46] Mazzaschi, G., Milanese, G., Pagano, P., Madeddu, D., Gnetti, L., Trentini, F., *et al.* (2020) Integrated CT Imaging and Tissue Immune Features Disclose a Radio-Immune Signature with High Prognostic Impact on Surgically Resected NSCLC. *Lung Cancer*, **144**, 30-39. <https://doi.org/10.1016/j.lungcan.2020.04.006>
- [47] Trentini, F., Mazzaschi, G., Milanese, G., Pavone, C., Madeddu, D., Gnetti, L., *et al.* (2021) Validation of a Radiomic Approach to Decipher NSCLC Immune Microenvironment in Surgically Resected Patients. *Tumori Journal*, **108**, 86-92. <https://doi.org/10.1177/03008916211000808>
- [48] Yoon, H.J., Kang, J., Park, H., Sohn, I., Lee, S. and Lee, H.Y. (2020) Deciphering the Tumor Microenvironment through Radiomics in Non-Small Cell Lung Cancer: Correlation with Immune Profiles. *PLOS ONE*, **15**, e0231227. <https://doi.org/10.1371/journal.pone.0231227>
- [49] Kim, H., Kwon, H.J., Han, Y.B., Park, S.Y., Kim, E.S., Kim, S.H., *et al.* (2019) Increased CD3⁺ T Cells with a Low FOXP3⁺/CD8⁺ T Cell Ratio Can Predict Anti-PD-1 Therapeutic Response in Non-Small Cell Lung Cancer Patients.

- Modern Pathology*, **32**, 367-375. <https://doi.org/10.1038/s41379-018-0142-3>
- [50] Li, H., Yu, H., Lan, S., Zhao, D., Liu, Y. and Cheng, Y. (2021) Aberrant Alteration of Circulating Lymphocyte Subsets in Small Cell Lung Cancer Patients Treated with Radiotherapy. *Technology in Cancer Research & Treatment*, **20**, 1-10. <https://doi.org/10.1177/15330338211039948>
- [51] Geng, Y., Shao, Y., He, W., Hu, W., Xu, Y., Chen, J., *et al.* (2015) Prognostic Role of Tumor-Infiltrating Lymphocytes in Lung Cancer: A Meta-analysis. *Cellular Physiology and Biochemistry*, **37**, 1560-1571. <https://doi.org/10.1159/000438523>
- [52] Hendry, S., Salgado, R., Gevaert, T., Russell, P.A., John, T., Thapa, B., *et al.* (2017) Assessing Tumor-Infiltrating Lymphocytes in Solid Tumors: A Practical Review for Pathologists and Proposal for a Standardized Method from the International Immuno-Oncology Biomarkers Working Group: Part 2: Tils in Melanoma, Gastrointestinal Tract Carcinomas, Non-Small Cell Lung Carcinoma and Mesothelioma, Endometrial and Ovarian Carcinomas, Squamous Cell Carcinoma of the Head and Neck, Genitourinary Carcinomas, and Primary Brain Tumors. *Advances in Anatomic Pathology*, **24**, 311-335. <https://doi.org/10.1097/pap.000000000000161>
- [53] Zeng, D., Yu, Y., Ou, Q., Li, X., Zhong, R., Xie, C., *et al.* (2016) Prognostic and Predictive Value of Tumor-Infiltrating Lymphocytes for Clinical Therapeutic Research in Patients with Non-Small Cell Lung Cancer. *Oncotarget*, **7**, 13765-13781. <https://doi.org/10.18632/oncotarget.7282>
- [54] Chen, L., Chen, L., Ni, H., Shen, L., Wei, J., Xia, Y., *et al.* (2023) Prediction of CD3 T Cells and CD8 T Cells Expression Levels in Non-Small Cell Lung Cancer Based on Radiomic Features of CT Images. *Frontiers in Oncology*, **13**, Article 1104316. <https://doi.org/10.3389/fonc.2023.1104316>
- [55] Patel, S.A., Nilsson, M.B., Yang, Y., Le, X., Tran, H.T., Elamin, Y.Y., *et al.* (2023) IL6 Mediates Suppression of T- and NK-Cell Function in EMT-Associated TKI-Resistant EGFR-Mutant NSCLC. *Clinical Cancer Research*, **29**, 1292-1304. <https://doi.org/10.1158/1078-0432.ccr-22-3379>
- [56] Chuang, T., Lai, W., Gabre, J.L., Lind, D.E., Umamathy, G., Bokhari, A.A., *et al.* (2023) ALK Fusion NSCLC Oncogenes Promote Survival and Inhibit NK Cell Responses via *SERPINB4* Expression. *Proceedings of the National Academy of Sciences of the United States of America*, **120**, e2216479120. <https://doi.org/10.1073/pnas.2216479120>
- [57] Meng, X., Xu, H., Liang, Y., Liang, M., Song, W., Zhou, B., *et al.* (2024) Enhanced CT-Based Radiomics Model to Predict Natural Killer Cell Infiltration and Clinical Prognosis in Non-Small Cell Lung Cancer. *Frontiers in Immunology*, **14**, Article 1334886. <https://doi.org/10.3389/fimmu.2023.1334886>
- [58] Hou, R., Xia, W., Zhang, C., Shao, Y., Zhu, X., Feng, W., *et al.* (2023) Dosiomics and Radiomics Improve the Prediction of Post-Radiotherapy Neutrophil-lymphocyte Ratio in Locally Advanced Non-Small Cell Lung Cancer. *Medical Physics*, **51**, 650-661. <https://doi.org/10.1002/mp.16829>
- [59] Weisberg, E.M., Chu, L.C., Park, S., Yuille, A.L., Kinzler, K.W., Vogelstein, B., *et al.* (2020) Deep Lessons Learned: Radiology, Oncology, Pathology, and Computer Science Experts Unite around Artificial Intelligence to Strive for Earlier Pancreatic Cancer Diagnosis. *Diagnostic and Interventional Imaging*, **101**, 111-115. <https://doi.org/10.1016/j.diii.2019.09.002>
- [60] Kang, J., Rancati, T., Lee, S., Oh, J.H., Kerns, S.L., Scott, J.G., *et al.* (2018) Machine Learning and Radiogenomics: Lessons Learned and Future Directions. *Frontiers in Oncology*, **8**, Article 228. <https://doi.org/10.3389/fonc.2018.00228>
- [61] Harding-Theobald, E., Louissaint, J., Maraj, B., Cuaresma, E., Townsend, W., Mendiratta-Lala, M., *et al.* (2021) Systematic Review: Radiomics for the Diagnosis and Prognosis of Hepatocellular Carcinoma. *Alimentary Pharmacology & Therapeutics*, **54**, 890-901. <https://doi.org/10.1111/apt.16563>
- [62] Duron, L., Savatovsky, J., Fournier, L. and Lecler, A. (2021) Can We Use Radiomics in Ultrasound Imaging? Impact of Preprocessing on Feature Repeatability. *Diagnostic and Interventional Imaging*, **102**, 659-667. <https://doi.org/10.1016/j.diii.2021.10.004>
- [63] Qi, Y., Zhao, T. and Han, M. (2022) The Application of Radiomics in Predicting Gene Mutations in Cancer. *European Radiology*, **32**, 4014-4024. <https://doi.org/10.1007/s00330-021-08520-6>