

肠道菌群 - 代谢物轴调控绝经后骨质疏松骨代谢失衡的机制研究进展

黄笑^{1,2}

¹广西中医药大学研究生院, 广西 南宁

²广西中医药大学第一附属医院内分泌科, 广西 南宁

收稿日期: 2026年4月16日; 录用日期: 2026年5月18日; 发布日期: 2026年5月27日

摘要

绝经后骨质疏松症是一种以骨量减少和骨微结构破坏为特征的全身性骨病, 其核心在于骨代谢失衡。近年来, 肠道菌群及其代谢产物构成的“肠道菌群-代谢物轴”在调控骨代谢中的作用日益凸显。本文综述了绝经后状态下肠道菌群紊乱的特征, 重点探讨了短链脂肪酸、胆汁酸、色氨酸代谢物等肠道菌群衍生代谢物如何通过影响免疫系统、肠屏障功能及内分泌信号等途径, 调控骨代谢失衡的分子机制, 旨在阐明该轴在疾病发病中的作用, 为发现新的诊断标志物和治疗靶点提供依据。

关键词

绝经后骨质疏松, 肠道菌群, 代谢物, 肠-骨轴, 骨代谢失衡, 免疫调节

Research Progress on the Mechanism of Gut Microbiota-Metabolite Axis in Regulating Bone Metabolic Imbalance in Postmenopausal Osteoporosis

Xiao Huang^{1,2}

¹Graduate School, Guangxi University of Chinese Medicine, Nanning Guangxi

²Department of Endocrinology, The First Affiliated Hospital of Guangxi University of Chinese Medicine, Nanning Guangxi

Received: April 16, 2026; accepted: May 18, 2026; published: May 27, 2026

Abstract

Postmenopausal osteoporosis is a systemic bone disease characterized by reduced bone mass and deteriorated bone microarchitecture, with the core pathogenesis lying in imbalanced bone metabolism. In recent years, the “gut microbiota-metabolite axis” composed of gut microbiota and their metabolites has played an increasingly prominent role in regulating bone metabolism. This article reviews the characteristics of gut microbiota dysbiosis in the postmenopausal state, and focuses on the molecular mechanisms by which gut microbiota-derived metabolites such as short-chain fatty acids, bile acids, and tryptophan metabolites regulate bone metabolic imbalance via modulating the immune system, intestinal barrier function, and endocrine signaling. It aims to clarify the role of this axis in the disease pathogenesis and provide evidence for identifying novel diagnostic biomarkers and therapeutic targets.

Keywords

Postmenopausal Osteoporosis, Gut Microbiota, Metabolites, Gut-Bone Axis, Bone Metabolic Imbalance, Immunoregulation

Copyright © 2026 by author(s) and Hans Publishers Inc.

This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

1. 引言

绝经后骨质疏松症是老年女性常见的骨骼疾病，雌激素缺乏是其公认的主要诱因[1]。其发病机制复杂，传统治疗如双膦酸盐、降钙素等虽能抑制骨吸收，但存在颌骨坏死、不典型骨折等严重并发症，因此亟待探索新的治疗靶点[2][3]。近年来研究发现，肠道菌群的结构、功能及代谢产物与宿主健康的关系密切。绝经后雌激素缺乏可导致肠道菌群失调，进而通过影响代谢物的产生，远程调控骨骼稳态，形成“肠道菌群-代谢物-骨”轴[1][3]。这一轴心涉及免疫调节、肠屏障功能、内分泌信号等多重通路，是连接雌激素缺乏与骨代谢失衡的关键桥梁[4]。

经临床与动物研究证实，绝经后骨质疏松患者或去卵巢模型动物的肠道菌群多样性降低，菌群结构发生显著改变，例如厚壁菌门与拟杆菌门的比例(F/B值)升高，而有益菌如乳酸杆菌属、双歧杆菌属的丰度下降[5][6]。这种菌群紊乱可导致肠道上皮屏障功能受损，表现为紧密连接蛋白(如 Occludin、ZO-1)表达下调，肠道通透性增加，形成“肠漏”[7]。肠屏障的破坏使得内毒素(如脂多糖 LPS)及有害代谢产物易于易位进入血液循环，引发系统性低度炎症[8]。炎症状态可激活免疫系统，特别是影响辅助性 T 细胞 17 与调节性 T 细胞之间的平衡，导致促炎细胞因子(如 TNF- α 、IL-6、IL-17)水平升高，而抗炎因子(如 IL-10、TGF- β)水平降低[4][9]。这些炎症因子可直接或间接刺激破骨细胞前体分化，增强骨吸收活性，同时抑制成骨细胞功能，从而加剧骨丢失[10]。

肠道菌群代谢产物，尤其是短链脂肪酸，在调控骨免疫和骨代谢中扮演着核心角色。SCFAs 主要由肠道中的膳食纤维经细菌发酵产生[9]。研究表明，绝经后骨质疏松状态下，产 SCFAs 的细菌丰度减少，导致粪便和血清中 SCFAs 水平下降[11][12]。SCFAs 不仅能为肠道上皮细胞提供能量、维护屏障完整性，还能作为信号分子调节免疫细胞功能[13]。此外，SCFAs 还能通过影响成骨细胞和破骨细胞内的信号通路(如 Wnt/ β -catenin、RANKL/OPG)直接调控骨代谢[12]。因此，肠道菌群及其代谢产物构成的网络，通过免疫、代谢和屏障等多重途径，深刻影响着绝经后骨质疏松的进程。

2. 绝经后骨质疏松症中肠道菌群的结构与功能变化

2.1. 雌激素缺乏诱导的肠道菌群生态失衡特征

在绝经后女性及卵巢切除(OVX)动物模型中, 肠道菌群的总体多样性显著降低, 菌群结构发生特异性改变。一项针对 106 名绝经后女性的研究发现, 与骨密度正常者相比, 骨质疏松患者的肠道细菌丰富度和多样性均有所下降[14]。在 OVX 大鼠模型中, 也观察到操作分类单元(OTUs)数量显著增加, 且 α 多样性(如 ACE、CHAO、Shannon 和 Simpson 指数)在卵巢切除后呈现上升趋势, 表明菌群结构发生了深刻变化[15]。具体表现为有益菌丰度下降, 而潜在致病菌或促炎菌丰度相对增加。例如, 在绝经后骨质疏松患者中, 有益菌如乳酸杆菌属(Lactobacillus)和双歧杆菌属(Bifidobacterium)的相对丰度可能降低[6]。相反, 一些与炎症相关的菌属, 如拟杆菌属(Bacteroides)的某些菌种和脱硫弧菌属(Desulfovibrio)的丰度可能增加[15]。这种生态失衡与肠道通透性增加、系统性低度炎症状态密切相关, 为骨代谢失衡创造了病理环境。研究表明, OVX 小鼠的肠道屏障功能被显著破坏, 导致肠道泄漏, 这被认为是引发系统性炎症和后续骨丢失的关键环节[16]。

2.2. 失调菌群的功能改变及其对宿主代谢的影响

失调的肠道菌群其碳水化合物发酵、胆汁酸代谢和氨基酸转化等关键代谢功能发生紊乱[15]。功能基因分析显示, 与短链脂肪酸(SCFAs)合成相关的基因表达下调, 而与脂多糖(LPS)生物合成相关的基因可能上调。例如, 在 OVX 大鼠的早期骨质疏松阶段, 涉及碳水化合物合成与代谢的肠道菌群功能基因有所增加, 这可能反映了菌群对宿主代谢紊乱的适应性改变, 但同时也可能加剧代谢失衡[15]。这些功能改变直接影响了肠道代谢产物的谱系和浓度, 是菌群调控骨代谢的物质基础。研究表明, OVX 小鼠肠道和血清中的 SCFAs 水平显著降低, 而 LPS 等促炎代谢物可能增加[17]。这种代谢谱的改变, 特别是 SCFAs 的减少, 会削弱其对骨形成的促进作用和对破骨细胞生成的抑制作用[17]。同时, 胆汁酸代谢的紊乱也被证实与绝经后骨质疏松的发生发展相关[18]。因此, 肠道菌群的功能性失调通过改变关键代谢产物的平衡, 深刻影响着宿主的骨代谢稳态。

3. 关键肠道菌群代谢物对骨代谢的调控机制

3.1. 短链脂肪酸的免疫调节与直接成骨作用

短链脂肪酸(SCFAs)是膳食纤维经肠道菌群发酵的主要产物。研究表明, 绝经后骨质疏松(PMOP)患者的肠道菌群发生显著改变, 其粪便中 SCFAs 水平常降低[14]。这种 SCFAs 的减少与骨代谢失衡密切相关。SCFAs, 尤其是丁酸, 在骨代谢调控中扮演着核心角色。一方面, 丁酸能通过免疫调节途径影响骨稳态, 例如促进调节性 T 细胞(Tregs)的分化和功能, 同时抑制辅助性 T 细胞 17(Th17)的活性, 从而下调促破骨细胞因子的产生, 从免疫层面抑制过度的骨吸收[19]。另一方面, 丁酸作为组蛋白去乙酰化酶抑制剂, 能直接作用于成骨前体细胞, 上调成骨相关基因(如 Runx2)的表达, 促进骨形成[20]。此外, SCFAs 还能通过激活 G 蛋白耦联受体(如 GPR43)影响骨细胞信号传导。动物实验也证实, 补充能增加丁酸水平的益生元或中药, 可通过提升 SCFAs 水平改善卵巢切除(OVX)大鼠的骨微结构和骨密度[21] [22]。

3.2. 胆汁酸代谢重编程及其信号通路干预

绝经后胆汁酸池大小和组成变化, 次级胆汁酸(如脱氧胆酸)比例可能因肠道菌群结构改变而波动[23]。胆汁酸作为重要肠道菌群代谢物, 可通过激活法尼醇 X 受体(FXR)和 G 蛋白耦联胆汁酸受体 1 (TGR5)等关键通路干预骨代谢。激活骨细胞 FXR 可能抑制成骨, 而激活免疫细胞(如巨噬细胞) TGR5 则具有抗炎

效应,可能间接保护骨量[24]。特定胆汁酸及其代谢重编程在骨质疏松中显示保护潜力,例如在去势(OVX)骨质疏松大鼠模型中,罗非鱼头脂质干预可通过调节肠道菌群影响初级胆汁酸代谢改善骨丢失[25]。此外,中药淫羊藿苷治疗 OVX 大鼠后,粪便代谢物中胆汁酸水平变化与改善的骨微结构参数相关[26]。这些研究表明,绝经后胆汁酸代谢重编程及其与肠道菌群的相互作用是影响骨稳态的重要机制。

3.3. 色氨酸代谢物与芳烃受体信号轴

肠道菌群能将膳食色氨酸代谢为吲哚等生物活性物质[27],这些物质是芳烃受体(AhR)的内源性配体。AhR 信号在骨代谢中作用复杂:在破骨细胞前体细胞中,其激活抑制分化从而限制骨吸收[27];在成骨细胞中,适度激活可能有益,过度激活则可能不利。绝经后雌激素缺乏可能导致肠道菌群失调,影响色氨酸代谢并改变 AhR 配体水平[28],这可能解除对破骨前体细胞的抑制,加剧骨吸收。研究证实绝经后骨质疏松女性肠道菌群组成及功能(如氨基酸代谢通路)与健康女性存在差异[28]。现有证据强烈提示,肠道菌群来源的色氨酸代谢物通过 AhR 信号通路,是连接肠道环境与骨代谢平衡的一个潜在关键节点。

4. 靶向肠道菌群 - 代谢物轴的干预策略与展望

4.1. 益生菌、益生元与合生元的应用研究

益生菌、益生元及合生元作为调控“肠道菌群 - 代谢物轴”的手段,在绝经后骨质疏松(PMOP)防治中展现出潜力。特定益生菌株如乳杆菌属和双歧杆菌属能改善骨代谢。动物实验表明,口服特定乳杆菌来源的肽聚糖(PGNs),特别是植物乳杆菌(*Lactiplantibacillus plantarum*)的 PGN,能恢复卵巢切除(OVX)小鼠的骨小梁体积,其机制涉及激活 NOD2 信号通路、促进成骨细胞分化并抑制破骨细胞分化,同时调节肠道菌群组成[29]。此外,益生菌如鼠李糖乳杆菌(*Lactocaseibacillus rhamnosus*)及其益生元,可通过调节肠道菌群、增加短链脂肪酸(SCFAs)产量并增强抗炎反应,从而减轻高脂饮食诱导的肥胖及相关骨代谢紊乱[30]。益生元如菊粉型果糖和低聚半乳糖,能促进双歧杆菌和乳杆菌等有益菌的生长,增加 SCFAs 的产量,这些代谢物具有抗炎特性并能增强免疫调节,从而对骨骼健康产生积极影响[31]。合生元即益生菌与益生元的组合,可能产生协同效应。例如,将益生菌与富含多酚的葡萄籽粉等益生元结合使用,能更有效地改善高脂饮食诱导的肠道菌群失调和肥胖[32]。尽管临床前研究结果令人鼓舞,但这些干预措施用于 PMOP 治疗的有效性、最佳菌株选择及剂量仍需大规模临床试验进一步验证[1]。

4.2. 饮食模式与菌群调控

饮食是塑造肠道菌群的基础策略,特定饮食模式通过调控“肠道菌群 - 代谢物轴”影响绝经后骨骼健康。地中海饮食等富含纤维的模式能塑造健康菌群,促进短链脂肪酸(SCFAs)生成。研究表明,SCFAs 作为关键代谢物,对维持肠道稳态、支持免疫、增强屏障及减轻炎症至关重要[33],这些作用与更高骨密度和更低骨折风险相关。临床研究发现,绝经后骨质疏松患者饮食有特征性改变,如摄入更多红肉和酒精,而健康对照组摄入更多酸奶、水果和茶[34]。限制高脂饮食可防止绝经后菌群失调和炎症,从而保护骨骼。在高脂饮食诱导的代谢综合征模型中,补充 SCFAs (如丁酸、丙酸和乙酸)能改善代谢参数、减轻肝脏损伤,并通过调节氧化应激发挥有益作用,其机制涉及 SCFA 受体(GPR41, GPR43, GPR109)和肠道菌群改变[35]。此外,富含多酚和 Omega-3 脂肪酸的饮食,如地中海饮食和生酮饮食,能促进菌群平衡、降低全身炎症,增强神经和骨骼韧性[36]。因此,通过调整整体饮食结构来优化肠道菌群及其代谢产物,是预防和管理绝经后骨质疏松的基础策略之一。

4.3. 粪菌移植与后生元疗法前景

粪菌移植(FMT)与后生元疗法是靶向“肠道菌群 - 代谢物轴”的前沿干预策略,为绝经后骨质疏松治

疗提供了新思路。FMT 通过移植健康供体的完整微生物群落直接重塑肠道生态,在动物模型中已证明可改善骨微结构,显示了肠道菌群的治病潜力[5]。然而, FMT 在人体应用仍面临供体筛选、安全性及伦理等挑战。相比之下, 后生元(如益生菌的无生命成分或其代谢产物)可能提供更安全、精准的干预方式,能绕过活菌定植的不确定性。例如,短链脂肪酸(SCFAs)作为重要后生元,在心血管疾病管理中显示出潜力,通过作用于 GPR41 等受体调控血压等过程[37]。在 PMOP 研究中, 补充 SCFAs 能通过调节受体和改变菌群,对肝脏代谢和炎症产生有益影响[38]。此外, 特定细菌成分如胞外多糖(EPS)也被视为后生元,能增加胆汁酸排泄和 SCFA 产量,改善脂质代谢和肠道健康[39]。这些发现表明, 后生元疗法是未来极具潜力的方向,为实现精准干预提供了可能。

5. 结论

综上所述, 绝经后骨质疏松的病理机制远非单纯的雌激素缺乏,其核心在于由此触发的“肠道菌群-代谢物轴”系统性失衡。本综述系统阐明,以短链脂肪酸、胆汁酸、色氨酸代谢物为代表的肠道菌群衍生代谢物,构成了连接肠道与骨骼的化学信使网络。它们通过调节免疫细胞功能、影响内分泌信号(如胰岛素样生长因子-1、5-羟色胺)以及维护肠道屏障完整性等多条并行且交互的通路,精密调控成骨细胞与破骨细胞的活性平衡。这一轴心的揭示,将骨骼健康的研究视角从局部骨微环境拓展至全身性的代谢与免疫稳态,为理解绝经后骨质疏松提供了全新的整合性框架。

在平衡不同观点时,需认识该领域的复杂性。尽管临床前研究支持特定益生菌或代谢物的骨保护作用,但人类试验结果存在异质性,可能源于遗传背景、菌群状态、饮食和干预差异。例如,一些研究强调了丁酸盐,另一些突出胆汁酸或吡啶类代谢物,这揭示了“肠道菌群-代谢物轴”的冗余性和个体特异性。未来研究应致力于绘制不同代谢物网络如何动态协同以维持骨平衡的全局图谱。

展望未来,靶向“肠道菌群-代谢物-骨”轴为绝经后骨质疏松的防治开辟了极具前景的新途径。这包括精准化的益生菌/元疗法、个性化的饮食营养调整以及粪菌移植等新兴微生物组疗法。然而,要将潜力转化为普适的临床实践,后续研究必须向纵深发展:一是需要利用多组学技术,更精细地解析驱动骨保护或骨破坏的具体关键菌种、代谢产物及其下游细胞分子靶点;二是必须设计和开展大规模、长周期、随机对照的临床研究,以确证这些干预措施在真实世界中的长期疗效、安全性以及最佳应用方案。最终,通过整合传统抗骨质疏松治疗与新兴的微生态调控策略,有望为绝经后女性构建起更全面、个性化的骨骼健康管理新范式。

参考文献

- [1] 罗建周, 吴太林, 段春光, 等. 绝经后骨质疏松免疫防治新靶点: 肠道菌群[J]. 中华骨与关节外科杂志, 2021, 14(8): 727-732. <https://clmd.academy/P7JEZBK8D/>
- [2] 李丹, 赵新正, 于金晟, 等. 基于肠道菌群防治绝经后骨质疏松症的相关研究进展[J]. 浙江临床医学, 2024, 26(1): 147-150. <https://clmd.academy/PNGZZTGD3/>
- [3] Xu, Q., Li, D., Chen, J., Yang, J., Yan, J., Xia, Y., et al. (2022) Crosstalk between the Gut Microbiota and Postmenopausal Osteoporosis: Mechanisms and Applications. *International Immunopharmacology*, **110**, Article ID: 108998. <https://doi.org/10.1016/j.intimp.2022.108998>
- [4] Qi, P., Xie, R., Liu, H., Zhang, Z., Cheng, Y., Ma, J., et al. (2024) Mechanisms of Gut Homeostasis Regulating Th17/Treg Cell Balance in PMOP. *Frontiers in Immunology*, **15**, Article 1497311. <https://doi.org/10.3389/fimmu.2024.1497311>
- [5] Guan, Z., Xuanqi, Z., Zhu, J., Yuan, W., Jia, J., Zhang, C., et al. (2023) Estrogen Deficiency Induces Bone Loss through the Gut Microbiota. *Pharmacological Research*, **196**, Article ID: 106930. <https://doi.org/10.1016/j.phrs.2023.106930>
- [6] 王颖颖, 邹丽莎, 金海泉. 绝经后骨质疏松患者骨免疫系统相关因子与肠道菌群的相关性[J]. 临床误诊误治, 2022, 35(1): 70-74. <https://clmd.academy/P4RPVPCDY/>
- [7] Zhang, Z., Lin, T., Meng, Y., Hu, M., Shu, L., Jiang, H., et al. (2021) FOS/GOS Attenuates High-Fat Diet Induced Bone

- Loss via Reversing Microbiota Dysbiosis, High Intestinal Permeability and Systemic Inflammation in Mice. *Metabolism*, **119**, Article ID: 154767. <https://doi.org/10.1016/j.metabol.2021.154767>
- [8] Zhang, Y., Cao, M., Li, Y., Lu, P., Dai, G., Zhang, M., *et al.* (2022) Fecal Microbiota Transplantation Ameliorates Bone Loss in Mice with Ovariectomy-Induced Osteoporosis via Modulating Gut Microbiota and Metabolic Function. *Journal of Orthopaedic Translation*, **37**, 46-60. <https://doi.org/10.1016/j.jot.2022.08.003>
- [9] Chen, Y., Xie, Y. and Yu, X. (2025) Progress of Research on the Gut Microbiome and Its Metabolite Short-Chain Fatty Acids in Postmenopausal Osteoporosis: A Literature Review. *Frontiers of Medicine*, **19**, 474-492. <https://doi.org/10.1007/s11684-025-1129-3>
- [10] Yao, Y., Cai, X., Chen, Y., Zhang, M. and Zheng, C. (2025) Estrogen Deficiency-Mediated Osteoimmunity in Postmenopausal Osteoporosis. *Medicinal Research Reviews*, **45**, 561-575. <https://doi.org/10.1002/med.22081>
- [11] Huang, K., Lin, C., Chuang, P., Yang, T., Tsai, Y., Li, Y., *et al.* (2025) Microbiota Diversity and Its Influence on Diabetic Osteoporosis Development. *Biochemical and Biophysical Research Communications*, **790**, Article ID: 152884. <https://doi.org/10.1016/j.bbrc.2025.152884>
- [12] Chen, Z., Liu, H., Chen, Y., Tang, Y., Tang, Y., Sarmiento, B., *et al.* (2025) Self-Replenishable Metabolically Augmented Synbiotic Microspheres Remodel Gut-Bone Homeostasis. *Advanced Materials*, **37**, e2500746. <https://doi.org/10.1002/adma.202500746>
- [13] Zhang, H., Qin, S., Zhang, X., Du, P., Zhu, Y., Huang, Y., *et al.* (2022) Dietary Resistant Starch Alleviates *Escherichia coli*-Induced Bone Loss in Meat Ducks by Promoting Short-Chain Fatty Acid Production and Inhibiting Malt1/NF- κ B Inflammasome Activation. *Journal of Animal Science and Biotechnology*, **13**, Article No. 92. <https://doi.org/10.1186/s40104-022-00739-7>
- [14] He, J., Xu, S., Zhang, B., Xiao, C., Chen, Z., Si, F., *et al.* (2020) Gut Microbiota and Metabolite Alterations Associated with Reduced Bone Mineral Density or Bone Metabolic Indexes in Postmenopausal Osteoporosis. *Aging*, **12**, 8583-8604. <https://doi.org/10.18632/aging.103168>
- [15] Ma, S., Qin, J., Hao, Y., Shi, Y. and Fu, L. (2020) Structural and Functional Changes of Gut Microbiota in Ovariectomized Rats and Their Correlations with Altered Bone Mass. *Aging*, **12**, 10736-10753. <https://doi.org/10.18632/aging.103290>
- [16] Chen, C., Lei, H., Zhao, Y., Hou, Y., Zheng, H., Zhang, C., *et al.* (2023) A Novel Small Molecule Effectively Ameliorates Estrogen Deficiency-Induced Osteoporosis by Targeting the Gut-Bone Signaling Axis. *European Journal of Pharmacology*, **954**, Article ID: 175868. <https://doi.org/10.1016/j.ejphar.2023.175868>
- [17] Chen, X., Zhang, Z., Hu, Y., Cui, J., Zhi, X., Li, X., *et al.* (2020) Lactulose Suppresses Osteoclastogenesis and Ameliorates Estrogen Deficiency-Induced Bone Loss in Mice. *Aging and disease*, **11**, 629-641. <https://doi.org/10.14336/ad.2019.0613>
- [18] Wen, K., Tao, L., Tao, Z., Meng, Y., Zhou, S., Chen, J., *et al.* (2020) Fecal and Serum Metabolomic Signatures and Microbial Community Profiling of Postmenopausal Osteoporosis Mice Model. *Frontiers in Cellular and Infection Microbiology*, **10**, Article 535310. <https://doi.org/10.3389/fcimb.2020.535310>
- [19] Sun, P., Zhang, C., Huang, Y., Yang, J., Zhou, F., Zeng, J., *et al.* (2022) Jiangu Granule Ameliorated OVX Rats Bone Loss by Modulating Gut Microbiota-SCFAs-Treg/Th17 Axis. *Biomedicine & Pharmacotherapy*, **150**, Article ID: 112975. <https://doi.org/10.1016/j.biopha.2022.112975>
- [20] Li, J., Yu, M., Pal, S., Tyagi, A.M., Dar, H., Adams, J., *et al.* (2020) Parathyroid Hormone-Dependent Bone Formation Requires Butyrate Production by Intestinal Microbiota. *Journal of Clinical Investigation*, **130**, 1767-1781. <https://doi.org/10.1172/jci133473>
- [21] Porwal, K., Pal, S., Kulkarni, C., Singh, P., Sharma, S., Singh, P., *et al.* (2020) A Prebiotic, Short-Chain Fructo-Oligosaccharides Promotes Peak Bone Mass and Maintains Bone Mass in Ovariectomized Rats by an Osteogenic Mechanism. *Biomedicine & Pharmacotherapy*, **129**, Article ID: 110448. <https://doi.org/10.1016/j.biopha.2020.110448>
- [22] 李伟举, 陈晓聪, 曾家莹, 等. 参骨颗粒通过肠道菌群改善骨质疏松的研究[J]. 中国骨质疏松杂志, 2024, 30(9): 1341-1347, 1352. <https://clmd.academy/PT58UJTAA/>
- [23] Bellissimo, M.P., Roberts, J.L., Jones, D.P., Liu, K.H., Taibl, K.R., Uppal, K., *et al.* (2020) Metabolomic Associations with Serum Bone Turnover Markers. *Nutrients*, **12**, Article 3161. <https://doi.org/10.3390/nu12103161>
- [24] Kwon, Y., Park, C., Lee, J., Park, D.H., Jeong, S., Yun, C., *et al.* (2021) Regulation of Bone Cell Differentiation and Activation by Microbe-Associated Molecular Patterns. *International Journal of Molecular Sciences*, **22**, Article 5805. <https://doi.org/10.3390/ijms22115805>
- [25] Zhu, Y., Liu, S., Mei, F., Zhao, M., Xia, G. and Shen, X. (2022) Tilapia Nilotica Head Lipids Improved Bone Loss by Regulating Inflammation and Serum Metabolism through Gut Microbiota in Ovariectomized Rats. *Frontiers in Nutrition*, **8**, Article 792793. <https://doi.org/10.3389/fnut.2021.792793>
- [26] Wang, S., Wang, S., Wang, X., Xu, Y., Zhang, X., Han, Y., *et al.* (2022) Effects of Icariin on Modulating Gut Microbiota

- and Regulating Metabolite Alterations to Prevent Bone Loss in Ovariectomized Rat Model. *Frontiers in Endocrinology*, **13**, Article 874849. <https://doi.org/10.3389/fendo.2022.874849>
- [27] Langan, D., Perkins, D.J., Vogel, S.N. and Moudgil, K.D. (2021) Microbiota-Derived Metabolites, Indole-3-Aldehyde and Indole-3-Acetic Acid, Differentially Modulate Innate Cytokines and Stromal Remodeling Processes Associated with Autoimmune Arthritis. *International Journal of Molecular Sciences*, **22**, Article 2017. <https://doi.org/10.3390/ijms22042017>
- [28] Rettedal, E.A., Ilesanmi-Oyelere, B.L., Roy, N.C., Coad, J. and Kruger, M.C. (2021) The Gut Microbiome Is Altered in Postmenopausal Women with Osteoporosis and Osteopenia. *JBMR Plus*, **5**, e10452. <https://doi.org/10.1002/jbm4.10452>
- [29] Kim, J., Park, O., Park, C., Kwon, Y., Yun, C. and Han, S.H. (2026) Oral Delivery of NOD2-Activating *Lactobacillus* Peptidoglycans Prevents Postmenopausal Osteoporosis via Gut Microbiota Modulation. *Biomedicine & Pharmacotherapy*, **195**, Article ID: 118976. <https://doi.org/10.1016/j.biopha.2026.118976>
- [30] Feng, X., Li, H., Tian, J., Han, X., Liang, W., Zhong, F., *et al.* (2025) Postbiotics from *Lactocaseibacillus rhamnosus* IOB820 Combat Obesity in HFD Mice by Modulating Gut Microbiota and Enhancing SCFA Production. *Nutrients*, **17**, Article 3525. <https://doi.org/10.3390/nu17223525>
- [31] Baheti, R., Deshkar, S., Jadhav, S., Mule, K., Jha, A., Giram, P., *et al.* (2026) Interplay of Probiotics, Prebiotics, Synbiotics and Postbiotics: A Review of Their Therapeutic Potential for Gastrointestinal Inflammation. *Food Research International*, **230**, Article ID: 118598. <https://doi.org/10.1016/j.foodres.2026.118598>
- [32] Seo, K., Gyu Lee, H., Young Eor, J., Jin Jeon, H., Yokoyama, W. and Kim, H. (2022) Effects of Kefir Lactic Acid Bacteria-Derived Postbiotic Components on High Fat Diet-Induced Gut Microbiota and Obesity. *Food Research International*, **157**, Article ID: 111445. <https://doi.org/10.1016/j.foodres.2022.111445>
- [33] Kukaev, E.N., Tokareva, A.O., Krogh-Jensen, O.A., Lenyushkina, A.A. and Starodubtseva, N.L. (2025) Gut Microbiota and Short-Chain Fatty Acids in the Pathogenesis of Necrotizing Enterocolitis in Very Preterm Infants. *Acta Naturae*, **17**, 38-51. <https://doi.org/10.32607/actanaturae.27623>
- [34] Chen, T., Meng, F., Wang, N., Hao, Y. and Fu, L. (2024) The Characteristics of Gut Microbiota and Its Relation with Diet in Postmenopausal Osteoporosis. *Calcified Tissue International*, **115**, 393-404. <https://doi.org/10.1007/s00223-024-01260-x>
- [35] Sharma, T., Ranawat, P., Garg, A., Rastogi, P. and Kaushal, N. (2025) Short-Chain Fatty Acids as a Novel Intervention for High-Fat Diet-Induced Metabolic Syndrome. *Molecular and Cellular Biochemistry*, **480**, 3169-3184. <https://doi.org/10.1007/s11010-024-05185-9>
- [36] Radhakrishnan, P., Viswanathan, K., Lini, S., Chinta, S. and Muthusamy, S. (2025) Unveiling Role of Gut Microbiota in Alzheimer's Disease: Mechanisms, Challenges and Future Perspectives. *Current Alzheimer Research*, **22**, 711-725. <https://doi.org/10.2174/0115672050403066250904112611>
- [37] Mousavi Ghahfarrokhi, S.S., Mohamadzadeh, M., Samadi, N., Fazeli, M.R., Khaki, S., Khameneh, B., *et al.* (2024) Management of Cardiovascular Diseases by Short-Chain Fatty Acid Postbiotics. *Current Nutrition Reports*, **13**, 294-313. <https://doi.org/10.1007/s13668-024-00531-1>
- [38] Mandaliya, D.K., Patel, S. and Seshadri, S. (2025) Postbiotic Potential of SCFAs on Metaflammation and Gut Microbiota Alteration in Diabetes. *Journal of Biosciences*, **50**, Article No. 57. <https://doi.org/10.1007/s12038-025-00531-5>
- [39] Ismael, M., Wu, J., Yang, H. and Zhong, Q. (2026) Dietary Interventions with *Schleiferilactobacillus harbinensis* Z171, Its EPS and Postbiotics Ameliorate Cholesterol Metabolism via Modulating the Gut-Liver Axis in High-Fat Diet-Fed Mice. *Molecular Nutrition & Food Research*, **70**, e70410. <https://doi.org/10.1002/mnfr.70410>